

## Inactivation of *Escherichia coli* O157:H7, *Salmonella enteritidis* and *Listeria monocytogenes* on the surface of tomatoes by neutral electrolyzed water

M.A. Deza, M. Araujo and M.J. Garrido

Institute of Food Research and Analysis, University of Santiago de Compostela,  
Santiago de Compostela, Spain

2003/0553: received 24 June 2003, revised 22 September 2003 and accepted 30 September 2003

### ABSTRACT

M.A. DEZA, M. ARAUJO AND M.J. GARRIDO. 2003.

**Aims:** To determine the efficacy of neutral electrolyzed water (NEW) in killing *Escherichia coli* O157:H7, *Salmonella enteritidis* and *Listeria monocytogenes*, as well as non-pathogenic *E. coli*, on the surface of tomatoes, and to evaluate the effect of rinsing with NEW on the organoleptic characteristics of the tomatoes.

**Methods and Results:** The bactericidal activity of NEW, containing 444 or 89 mg<sup>l</sup><sup>-1</sup> of active chlorine, was evaluated over pure cultures (8.5log CFU ml<sup>-1</sup>) of the above-mentioned strains. All of them were reduced by more than 6 log CFU ml<sup>-1</sup> within 5 min of exposure to NEW. Fresh tomatoes were surface-inoculated with the same strains, and rinsed in NEW (89 mg<sup>l</sup><sup>-1</sup> of active chlorine) or in deionized sterile water (control), for 30 or 60 s. In the NEW treatments, independent of the strain and of the treatment time, an initial surface population of about 5 log CFU sq.cm<sup>-1</sup> was reduced to 5log CFU sq.cm<sup>-1</sup>, and no cells were detected in the washing solution by plating procedure. A sensory evaluation was conducted to ascertain possible alterations in organoleptic qualities, yielding no significant differences with regard to untreated tomatoes.

**Significance and Impact of the Study:** Rinsing in NEW reveals as an effective method to control the presence of *E. coli* O157:H7, *S. enteritidis* and *L. monocytogenes* on the surface of fresh tomatoes, without affecting their organoleptic characteristics. This indicates its potential application for the decontamination of fresh produce surfaces.

**Keywords:** ANK-Anolyte, disinfectant, *E. coli* O 157 :H7 , *L. monocytogenes*, neutral electrolyzed water, organoleptic quality, rinsing fresh tomatoes, *S. enteritidis*.

### INTRODUCTION

Fruits and vegetables can become contaminated with pathogenic micro-organisms while growing in fields, during harvesting and postharvest handling, processing and distribution (Beuchat 1996). Human gastroenteritis has been epidemiologically linked to the consumption of ready-to-eat salads contaminated with enterotoxigenic *Escherichia coli* (Abdul-Raouf et al. 1993) and *Listeria monocytogenes* (Beuchat and Brackett 1991 ); outbreaks of salmonellosis have been attributed to the consumption of contaminated tomatoes (Zhuang et al. 1995; Beuchat 1996). Also, the growth of *L. monocytogenes* and *Salmonella spp.* on the surface of whole fresh-cut tomatoes has been reported (Asplund and Nurmi 1991; Beuchat and Brackett 1991).

*Correspondence to:* M..A. Deza, Instituto de Investigacion y Analisis alimentarios, Universidad de Santiago de Compostela, Campus Sur, E-15782 Santiago de Compostela, Spain ( e-mail: madez@usc.es)

Washing fresh produce with running tap water may remove soil and other debris, but it has a limited effect on surface micro-organisms that occur at populations ranging from  $10^3$  to  $10^9$  CFU  $g^{-1}$  (Oseki et al. 2001). A variety of disinfectants (chlorine, hydrogen peroxide, organic acids, ozone, etc.) have been used to reduce the bacterial population on fruits and vegetables. However, besides their potential toxicity, they cannot completely remove or inactivate micro-organisms on fresh produce (Koseki and Itoh 2001; Park et al. 2001).

In recent years, acidic electrolyzed water (AEW) and neutral electrolyzed water (NEW) have been introduced for application as sanitizers. These solutions are generated by electrolysis of a dilute NaCl solution passing through the anode of a membrane electrolyser. AEW has a strong bactericidal effect on most known pathogenic bacteria due to its low pH (2-4) and high oxidation-reduction potential (ORP > 1000 mV), and because it contains active oxidizers like hypochlorous acid (Kim et al. 2000b; Len et al. 2000), it is effective in killing food-borne pathogens in vitro conditions (Venkitanarayanan et al. 1999b; Kim et al. 2000a) and in reducing microbial counts and pathogens in vegetables (Koseki et al. 2001; Koseki and Itoh 2001; Park et al. 2001; Bari et al. 2003; Kim et al. 2003).

NEW is generated like AEW, but a part of the product formed at the anode is redirected into the cathode chamber, thus increasing the content of ClO<sup>-</sup> ions.

Because of its neutral pH, NEW does not contribute as aggressively as AEW to the corrosion of processing equipment or irritation of hands, and is more stable as chlorine loss is significantly reduced at pH 6-9 (Rojas and Guevara 2000; Len et al. 2002). Izumi (1999) has evaluated the effect of NEW (pH 6.8 and 20 mg  $l^{-1}$  active chlorine) on total microbial count in fresh-cut vegetables, obtaining reductions up to 2.6 log CFU  $g^{-1}$  without significant effect on tissue pH, surface colour and general appearance of vegetables.

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The aim of this work was to determine the effectiveness of NEW in killing *E. coli* O157:H7, *Salmonella enteritidis*, *L. monocytogenes* and nonpathogenic *E. coli*, in vitro and on the surface of tomatoes, with a view to its potential application to fresh produce and food contact surfaces as an antimicrobial treatment. A sensory evaluation was also performed in order to evaluate the effect of rinsing with NEW on the organoleptic characteristics of the tomatoes.

## MATERIALS AND METHODS

### Preparation of treatment solutions

NEW was generated using a **Eurostel unit (manufactured by Aquastel)**.

A 25% sodium chloride solution and tap water were simultaneously pumped into the generator to obtain amperage of  $32 \pm 2A$ . For this study, NEW (containing approx.  $444 \text{ mg l}^{-1}$  of active chlorine) was diluted 1 : 5 in deionized sterile water, to obtain a final active chlorine concentration of about  $89 \text{ mg l}^{-1}$ . Deionized sterile water was also used as control.

pH, ORP and active chlorine concentration were determined for both treatment solutions. The former magnitudes were measured after preparation, using a pH/ion/conductivity meter (CRISON micro-pH 2001) with a pH electrode (CRISON, 52-11) and an ORP electrode (CRISON platinum Ag/AgCl electrode, 52-61). The latter, by an iodometric method (APHA 1998).

### Treatment of pure culture

The strains used for this study were obtained from the Spanish Type Culture Collection (CECT): *E. coli* CECT 405 (ATCC strain 10536, proposed for testing antibiotics, antimicrobial preservatives and chemotherapeutic agents), *E. coli* O157:H7 CECT 4267 (ATCC strain 35150, isolated from an outbreak of haemorrhagic colitis, produces Shiga-like toxin I and II) *S. enteritidis* CECT 556 (isolated from water in Valencia, Spain) and *L. monocytogenes* CECT 4032 (isolated from soft cheese, associated with a case of meningitis). Strains were cultured on TSA plates [Tryptone Soy Broth (Panreac Quimica S.A., Barcelona, Spain) with the addition of 15 g l<sup>-1</sup> agar no.3 (Oxoid, Basingstoke, Hampshire, UK)] at 37°C for 24 h. The efficiency of NEW to produce a reduction in at least 5 logs in viable cell counts (bactericidal activity) in clean conditions was evaluated according to the European Standard UNE-EN 1276 (Anonymous 1998). One millilitre of bacterial culture of about 8.5 log CFU ml<sup>-1</sup> was transferred to sterile tubes together with 1 ml of sterile water. Eight millilitre of pure NEW (444 :1: 8.15 mg l<sup>-1</sup> active chlorine) or diluted 1 : 5 in deionized water (89 :1: 7.5 mg l<sup>-1</sup> active chlorine) were added. The tubes were hand-shaken to mix the resultant suspension, and incubated at room temperature (23 ± 2°C) for 5 min. Deionized water was used as a control.

Following treatment, 1 ml of each sample was transferred to 9 ml of neutralizing solution (sodium thiosulphate 0.5%) and the suspension hand shaken. After 5 min of neutralization, 1 ml of the appropriate dilution 1 : 10 in tryptone sodium chloride solution (pH 7.2 ± 0.2) was pour plated on TSA. The plates were incubated at 37 ± 1°C for 24 h. The experiment was repeated four times.

### Preparation and inoculation of tomatoes

Tomatoes (*Lycopersicon esculentum* var. Durinta) were purchased at a local supermarket and stored at 4°C, for a maximum of 3 days before testing. Units of similar size (70-80 g) without lesions on skin were used. Their surface area was calculated in order to obtain the number of CFU sq.cm<sup>-1</sup>. Tomatoes were first washed with tap water for 1 min and air-dried under sterile air in a laminar flow cabinet for 15 min in individual metallic strainers. For the inoculation of tomatoes, a bacterial suspension of 8.98-9.23 log CFU ml<sup>-1</sup> was prepared using 70 ml of tryptone sodium chloride solution. The bacterial population each inoculum was confirmed either by pouring 1 ml (for *E. coli*) or by surface-plating 0.1 ml (for *E. coli* O157:H7, *S. enteritidis* and *L. monocytogenes*) of appropriate dilutions of the suspension (using the same solution) on duplicate selective plates, using Coli ID medium (bioMerieux, Marcy l'Etoile, France) for *E. coli*, Sorbitol-MacConkey agar (Merck, Darmstadt, Denmark) for *E. coli* O157:H7, XLD agar (Oxoid) for *S. enteritidis*, and PALCAM agar (Merck) for *L. monocytogenes*. Plates of Coli ID, Sorbitol-MacConkey and XLD agar were incubated at 37°C for 24 h, and plates of PALCAM agar at 37°C for 48 h. Tomatoes were immersed for 1 min in the bacterial suspension of 9 log CFU ml<sup>-1</sup>, and then dried in individual sterile metallic strainers under sterile air in a laminar flow cabinet for 15 min at room temperature (23 ± 2°C).

### Treatment and bacteriological analysis of tomatoes

The initial population on tomato surface was obtained by swabbing the whole surface of an inoculated air-dried tomato with a sterile cotton swab moistened with 5 ml of sterile tryptone sodium chloride solution. Appropriate dilutions of this solution were plated onto selective plates as described above. Inoculated tomatoes were placed in individual sterile bags containing 100 ml of electrolyzed neutral water diluted 1 : 5, or sterile deionized water (control). The bags were shaken vigorously by hand for 30 or 60 s. After immersion in the treatment or control water, tomatoes were removed with a sterile metallic strainer and

allowed to drain completely. The whole surface of each tomato was then swabbed with a sterile cotton swab. The swab was washed in 5 ml of neutralizing solution and appropriate dilutions of this solution were plated onto selective plates. A volume of 1 ml of the treatment or control water was also transferred to 9 ml of neutralizing solution and appropriate dilutions were plated onto selective plates, as described in 'Preparation and inoculation of tomatoes'. All the experiments were conducted at room temperature ( $23 \pm 2^\circ\text{C}$ ), in order to imitate normal washing procedures for unprocessed produce at home.

### Sensory evaluation

The organoleptic properties of un-inoculated tomatoes treated with NEW (pure or diluted 1 : 5 in water) and untreated (washed with tap water) was evaluated by 12 panellists. Tomatoes were washed under tap water for 1 min, drained and submitted for 1 min to the above described treatment solutions, and air-dried for 6 h at  $23 \pm 2^\circ\text{C}$ . Panellists individually evaluated appearance, colour and taste of treated and untreated tomatoes. The quality evaluation was based on a five-point scale:

1, not acceptable; 2, limited quality; 3, normal; 4, good; 5, very good.

### Data analysis

All trials were repeated four times. Microbial counts were expressed as  $\log \text{CFU ml}^{-1}$  (washing solutions and inocula) or  $\text{CFU sq.cm}^{-1}$  (tomato surface). The reported values of plate count or physicochemical properties are the mean values over four individual trials  $\pm$  standard deviations.

Sensory evaluation values represent the mean of 12 values  $\pm$  standard deviations. Data were subjected to analysis of variance and Duncan's multiple range test using ST A TGRAPHICS (Statistical Graphics Corporation, Englewood Cliffs, NJ, USA). Significant differences in plate count data and in sensory evaluation were established by the least significant difference at the 0.05 level of significance.

## RESULTS

The pH, ORP and active chlorine concentration of treatment solutions used for each strain, are shown in Table 1

Table 1

### Physicochemical properties of tested solutions\*

Strain used in each treatment	Dionized water			NEW			NEW (diluted 1 :5 )		
	pH	ORP, mV	Cl, mg $l^{-1}$	pH	ORP, mV	Cl, mg $l^{-1}$	pH	ORP, mV	Cl, mg $l^{-1}$
<i>E. Coli</i>	6.01 $\pm$ 1.10	587 $\pm$ 9.0	0	8.13 $\pm$ 0.11	803 $\pm$ 11.0	430.6 $\pm$ 9.0	7.99 $\pm$ 0.21	750 $\pm$ 10.0	86.12 $\pm$ 7.2
<i>E.Coli O157:H7</i>	5.92 $\pm$ 0.56	551 $\pm$ 4.0	0	8.03 $\pm$ 0.23	816 $\pm$ 9.0	432 $\pm$ 5.1	8.15 $\pm$ 0.20	771 $\pm$ 7.0	86.40 $\pm$ 4.1
<i>S. enteritidis</i>	5.82 $\pm$ 0.23	575 $\pm$ 15.0	0	7.99 $\pm$ 0.15	795 $\pm$ 0.15	465 $\pm$ 7.5	8.19 $\pm$ 0.30	745 $\pm$ 8.0	93.00 $\pm$ 9.0
<i>L. monocytogenes</i>	6.30 $\pm$ 0.15	662 $\pm$ 9.0	0	8.20 $\pm$ 0.09	808 $\pm$ 7.5	450 $\pm$ 11.0	8.09 $\pm$ 0.05	760 $\pm$ 11.0	92.10 $\pm$ 10.0

\* Values are mean  $\pm$  S>D. of four repeated measurements

NEW, Neutral Electrolyzed water [ANK-Anolyte]

ORP, oxidation-reduction potential

Cl, active chlorine

All the strains treated for 5 min with NEW (containing 444 or 89 mg 1<sup>-1</sup> active chlorine) were reduced by more than 6 log CFU mg 1<sup>-1</sup>, as determined by plating procedure using the European Standard UNE-EN 1276 (Table 2). No reduction in bacterial counts was achieved in the control samples.

Table 2.

**Inactivation of *E. coli*, *E. coli* O157:H7, *S. enteritidis* and *L. monocytogenes* in pure culture by NEW (444 and 89mg 1<sup>-1</sup> active chlorine) in 5 min at 23 ± 2°C.**

Strain	Initial population (log CFU ml <sup>-1</sup> )	Surviving population after 5-min treatment (Log CFU 1 <sup>-1</sup> )		
		Control (dionized water)	NEW (444 ±8.15 mg 1 <sup>-1</sup> active chlorine)	NEW (dilution 1 : 5) (89 ± 7.5 mg 1 <sup>-1</sup> active chlorine)
<i>E. Coli</i>	7.51±0.11	7.50±0.20	<1	<1
<i>E.Coli</i> O157:H7	7.45±0.04	7.46±0.13	<1	<1
<i>S. enteritidis</i>	7.70±0.18	7.62±0.17	<1	<1
<i>L. monocytogenes</i>	7.51±0.17	7.53±0.21	<1	<1

Table 3 shows the inactivation of *E. coli*, *E. coli* O157:H7, *S. enteritidis* and *L. monocytogenes* on tomato surface treated with NEW. The initial population on tomato surface after inoculating and drying under cabinet for 15 min was between 5.29 and 5.58 log CFU sq .cm<sup>-1</sup>.

Washing with deionized water (control) reduced viable cells in all strains by approx. 2 log CFU sq.cm<sup>-1</sup> within 30 or 60 s.

Under treatment \with NEW, the populations on tomato surface of all strains were reduced by an average of 4.18 log CFU sq.cm<sup>-1</sup> in 30 s, and 4.74 log CFU sq.cm<sup>-1</sup> in 60 s. Populations of *E. coli* O157:H7, *S. enteritidis* and *L. monocytogenes* on the surface of tomatoes showed no significant difference between treatments with NEW at 30 or 60 s, whereas reduction in nonpathogenic *E. coli* population after treatment for 60 s was significantly lower ( $P \leq 0.05$ ) than after treatment for 30 s. Also, the populations of all strains, either after 30 s or after 60 s, were very similar , without significant strain dependence.

The surviving population in the wwashing solutions (NEW, diluted 1 : 5 or deionized water) is also indicated in Table 3.

Under treatment with NEW, no survivors were detected by plating procedure. In control water, an average of 5.35 log CFU ml<sup>-1</sup> was recovered.

No significant differences ( $P \leq 0,05$ ) were found in the sensory evaluation of uninoculated tomatoes washed with NEW (pure or diluted 1 : 5) or with tap water. On an ascending five-point scale, for both treated and control tomatoes, the mean values were between 3.21 and 3.54 for appearance, between 3-25 and 3.96 for smell, and between 3-08 and 3-83 for taste.